

## Immunofluorescence protocol for frozen mice & rat brain sections

Thaw	Thaw sections for 5 min at room temperature	5
Fixation	<ul style="list-style-type: none"> <li>Mark section surrounding with special "border" pen or nail polish</li> <li>Cover section either with 4% PFA (0.1M PB) for 6min at room temperature</li> </ul>	10
Wash	<ul style="list-style-type: none"> <li>Briefly rinse in PB</li> <li>Wash section 3x5 min in 0.1M PB on shaker platform</li> </ul>	15
Block	<ul style="list-style-type: none"> <li>BSA solution for 60 min, at room temperature             <ul style="list-style-type: none"> <li>3% Donkey serum, 1% BSA, 0.3% Tx, 0.1M PB</li> <li>Cover sections after adding serum and add wet paper to storage box to prevent dry out</li> </ul> </li> </ul>	60
Primary antibody	<ul style="list-style-type: none"> <li>Primary Anti-B in BSA solution, for 60min at room temperature             <ul style="list-style-type: none"> <li>Cover 1<sup>st</sup> half of the sections with primary antibody</li> <li>Cover 2<sup>nd</sup> half of section with BSA solution -&gt; control sections</li> </ul> </li> <li>Adding wet paper to the storage box and close with lid</li> </ul>	60
Wash	<ul style="list-style-type: none"> <li>Rinse &amp; Wash 3x5min in 0.1M PB on shaker platform</li> </ul>	8
Secondary antibody	<ul style="list-style-type: none"> <li>Secondary Anti-B in BSA solution for 60 min at room temperature             <ul style="list-style-type: none"> <li>Produce decided concentration: 1:1000                 <ul style="list-style-type: none"> <li>Add 2<sup>nd</sup> antibody to solution</li> <li>Cover all sections with solution</li> <li>Add wet paper and close lid in the dark</li> </ul> </li> </ul> </li> </ul>	60
Wash	<ul style="list-style-type: none"> <li>Rinse &amp; Wash 3x5min in 0.1 M PB on shaker platform in the dark</li> </ul>	10
Mount	<ul style="list-style-type: none"> <li>Dip slides in MilliQ water to prevent crystal formation</li> <li>Store slides in dark until dehydrated</li> <li>Mount with entellan neu, coverslip and store in dark at +4C</li> </ul>	120

### Solutions:

- use ca. 300 µl per glass slide (if not divided into sections) for each step

<ul style="list-style-type: none"> <li>- <b>0.1 M phosphate buffer</b> (2L, for one run per 25 glass slides)             <ul style="list-style-type: none"> <li>2 L MilliQ water</li> <li>+ 21.8 g Na<sub>2</sub>HPO<sub>4</sub></li> <li>+ 6.4 g NaH<sub>2</sub>PO<sub>4</sub></li> <li>Mix well until dissolved, adjust pH to 7.4 and store at +4C</li> </ul> </li> </ul>	<ul style="list-style-type: none"> <li>- <b>4% PFA in 0.1 M PB</b> <ul style="list-style-type: none"> <li>10 ml 0.1 M PB</li> <li>+ 400 mg Paraformaldehyde</li> <li>Heat to just under +58 C</li> <li>Add NaOH until clear</li> <li>Cool solution, adjust pH to 7.4</li> <li>Filter solution and store at +4C</li> </ul> </li> </ul>
<ul style="list-style-type: none"> <li>- <b>BSA solution (Blocking &amp; Incubation)</b> <ul style="list-style-type: none"> <li>10 ml 0.1 PB</li> <li>+ 300 µl donkey serum (3%)</li> <li>+ 100 mg BSA (1%)</li> <li>+ 30 µl Triton X 100 (0.3%)</li> <li>Mix well and store at +4C for 2-3 days</li> </ul> </li> </ul>	

